

Biotechnology Carbon Capture and Storage (CCS) by Mix-culture Green Microalgae to Enhancing Carbon Uptake Rate and Carbon Dioxide Removal Efficiency with Variation Aeration Rates in Closed System Photobioreactor

Astri Rinanti^{a*}, Kania Dewia^{a*}, Edwan Kardena^a, Dea Indriani Astuti^b

^aFaculty of Civil and Environmental Engineering, Bandung Institute of Technology, Indonesia. Jl. Ganesha No. 10, Bandung 40132

^bSchool of Life Sciences and Technology, Bandung Institute of Technology, Indonesia. Jl. Ganesha No. 10, Bandung 40132

*Corresponding author: astri@fun-dering.com

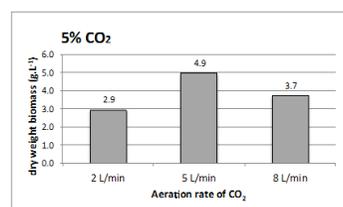
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Graphical abstract



Abstract

Carbon dioxide (CO₂) sequestration by green microalgae is receiving increased attention in alleviating the impact of increasing CO₂ in the atmosphere. The goal of this study was to explore the capacity of mixed culture green microalgae *Chlorella* sp, *Scenedesmus obliquus*, and *Ankistrodesmus* sp. as carbon capture and storage agent to enhance CO₂ uptake rate and CO₂ removal efficiency which was observed at elevated CO₂ aeration rates of 2, 5, and 8 L min⁻¹ supplied to vertical photobioreactor continuously in batch system culture. The operation condition of this research were 6.5-7.5 pH, temperature of 30°C, light intensity of 4000 lux with 16 hours light period and 8 hours dark period, and high pure CO₂ elevated level of 5 to 18 (concentration in %; v/v in the aeration gas) as inorganic carbon. The maximum CO₂ removal efficiency of the mix culture was 59.80% when the biomass was obtained at 4.90 gL⁻¹ and CO₂ flow rate (Lmin⁻¹) of 5 vvm in a vertical photobioreactor. The value of CO₂ removal efficiency improved by almost 200% and 120% as compared to that in the low and high aeration rate (2 Lmin⁻¹ and 8 Lmin⁻¹) respectively. The CO₂ up take rate of a mixed culture reach 979.62 mg carbon L⁻¹day⁻¹, which was enhancing by 3-fold in high aeration rate (8 Lmin⁻¹). The results showed that the CO₂ removal efficiency and carbon uptake rate was related to biomass concentration and aeration rate of CO₂ supplied.

Keywords : Carbon uptake rate; carbon dioxide removal efficiency; photobioreactor; microalgae

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1.0 INTRODUCTION

Carbon dioxide is usually emitted freely from industrial processes in an uncontrolled way. CO₂ concentration in the troposphere is getting serious attention as CO₂ is categorized as greenhouse gas that is believed to be the cause of global warming. Impacts of greenhouse gases are becoming more apparent mainly due to the increase of the earth's surface temperature [1-3]. Biological carbon capture and storage (CCS) technologies can be used to mitigate carbon emissions that would otherwise be released to the atmosphere. Research studies that utilized the potential of microalgae as CCS agent have been carried out in various countries, particularly in efforts towards adaptation and selection of microalgae species tolerant to high CO₂ concentrations and high CO₂ absorption rate. Most of the flue gases produced by most concentration make it advantageous for the microalgae to be the best candidate for creating a sustainable carbon sink. Since microalgae are producers, they have the ability to continuously undertake photosynthesis. One of the primary requirements for photosynthesis is atmospheric CO₂. Growing microalgae that

captures ambient CO₂ will remove carbon dioxide and sequester it in the form of biomass [4].

When CO₂ is injected in a culture, a concentration gradient builds up as it is consumed by cells and/or lost to the atmosphere. According to the two-film theory, mass transfer of CO₂ from the gas phase to the cell phase occurs through sequential stages [5-6]. The mass transfer of carbon dioxide from air into the growth media can be growth-limiting in dense microalgae cultures and its process photosynthesis. Milne *et al.* stated that the transfer of CO₂ from a gas to a liquid depends on many parameters [7]. Physical parameters such as gas aeration rate, CO₂ partial pressure, bubble diameter and lifetime can have large influences on the rate of transfer. Other studies have shown higher values of overall mass transfer coefficient are obtained at higher gas velocities [8]. An increase in superficial gas velocity causes an increase in gas holdup, which increases the interfacial area. There is an increase in interfacial area because higher gas velocity leads to higher momentum exchange between phases. As a result, bubbles break at a higher efficiency into smaller bubbles and the interfacial area becomes bigger.

The aim of the study was to observe the impact of different aeration rates of air-rich CO₂ to enhanced CO₂ removal efficiency and carbon uptake rate by mixed-culture green microalgae cultivated in vertical bubble photobioreactor.

2.0 MATERIAL AND METHODS

2.1 Mix-culture Microalgae and Artificial Growth Medium

The mix-culture green microalgae consisting of *Chlorella* sp., *Scenedesmus obliquus* and *Ankistrodesmus* sp. was originally isolated from the Bojong Soang wastewater treatment plant, Bandung, Indonesia. The microalgal was screened, and then a potential candidate was selected for Microbial Carbon Capture and Storage (MCCS) agent [9]. The microalgal cells were cultured in PHM (Phovasoli Haematococcus Media) artificial growth media [10].

2.2 Cultivation Microalgae in Vertical Photobioreactor and Experimental Condition

Vertical photobioreactor made of glass with a capacity of 10 L containing by 8 L PHM growth medium and an initial cell density of 10⁶ cell.ml⁻¹. The pure CO₂ gas supplied from the bottom of the photobioreactor with 5%, 10%, 15%, and 18% pure CO₂ level at different CO₂ aeration rate of 2.0, 5.0, and 8.0 L.min⁻¹, respectively, and temperature was adjusted at 30°C ± 1. CO₂ was injected from the bottom of the column to allow gas mixing with the medium. Sparger was attached at the bottom of the photobioreactor to convert the gas into small bubbles. Air is bubbled at the bottom. Microbubble sparging allows thorough mixing, CO₂ mass transfer and also removes O₂ produced during photo-synthesis. It was a strategy that provides good overall mixing, sufficient supply of CO₂, and efficient removal of O₂. Position 4 TL lamps uniformly placed outside the photobioreactor can be adjusted to obtain a light intensity of 4000 lux and light periods (light/dark; hour) of 16/8.

2.3 Measurement of Biomass Concentration and Growth rate of the Mix-culture Microalgae

Dry weight cell biomass of microalgae was obtained by evaporating the liquid in the cell culture. A total of 100 mL culture tube inserted into centrifuges, and then centrifuged at 3500 rpm for 10 minutes [11]. Supernatant was then removed from the tube pasta until just earned cells. Pasta cells were then put into a petri dish that had previously been weighed (x). Samples were put in the oven with a temperature of 105°C for one night to get a constant weight (y), and then stored in a desiccator for 30 minutes before re-weighed. Biomass (dry weight) to calculated by the formula: dry weight (X; mg) = y (mg) - x (mg). Specific growth rate (μ; d⁻¹) was calculated as follows [12]:

$$\mu = \frac{1}{X} \cdot \frac{dX}{dt} \quad (1)$$

2.4 Measurement of CO₂ Concentration and Determination of CO₂ Removal Efficiency

The CO₂ concentration in the influent gas and effluent gas was measured by Portable Combination Gas Detector RIKEN Model RX-515. Efficiency of CO₂ removal can be calculated by the following formula:

$$\frac{\text{Influent of CO}_2 - \text{Effluent of CO}_2}{\text{Influent of CO}_2} \times 100\% \quad (2)$$

An approximate formula (CO_{0.48}H_{1.83}N_{0.11}P_{0.01}) was used to make an expected estimate of the dry biomass yield [13] and carbon uptake rate was determined by using the following equation [14]:

$$\text{Carbon uptake rate} = C \times P \quad (3)$$

Where, C is the carbon content of the dry weight cell (g carbon.g biomass⁻¹), P is the productivity (g biomass.L⁻¹.d⁻¹). Results of elemental analysis in our study showed that the carbon content in the mix culture was 67.56%.

3.0 RESULTS AND DISCUSSION

3.1 Dry Weight of Biomass as Growth Response

Our previous study obtained the highest dry weight of biomass occurred from the culture which was supplied continuously with 5% (v/v) pure CO₂ [15]. Thus, the study of the impact of CO₂ aeration rates started with supplied 5% (v/v) pure CO₂.

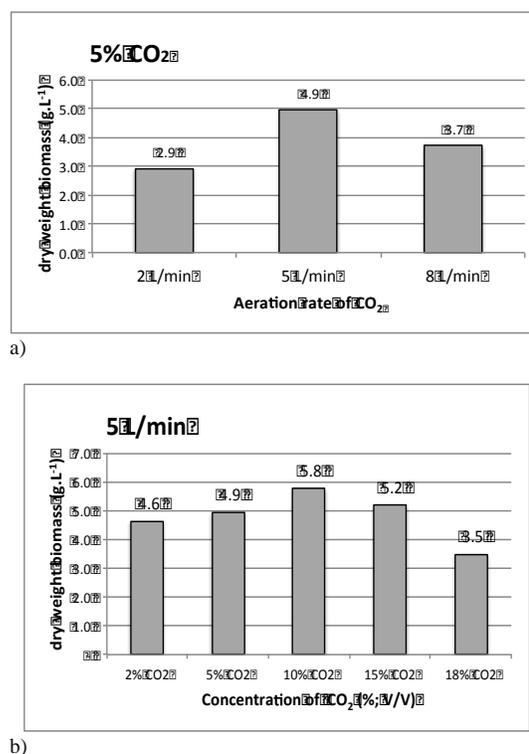


Figure 1 Dry weight biomass (a) with variation aeration rate of CO₂ and 5% concentration of CO₂, (b) with variation concentration of CO₂ (in %; v/v), CO₂ aeration rate of 5 L/min, all were supplied continuously

An increase in concentration of the CO₂ regardless of the flow rate resulted in a decrease in pH [16]. However, this study showed that aeration rate has a great influence on both growth and dry biomass yield because the growth medium has a weak buffering capacity. The pH drastically decreases when high level of CO₂ gas was supplied. It is possible that the low pH observed when pure CO₂ was used could have been the reason for the reduced growth rates. When dissolving in water, CO₂ equilibrates

into CO_2 (aq), HCO_3^- (aq), and CO_3^{2-} (aq). This lowers the pH, whereas at a pH of 6 and lower, CO_2 (aq) is dominant. At a pH of 6–9, HCO_3^- (aq) becomes more pronounced, and at a pH of 9 and above, CO_3^{2-} becomes predominant [16].

Another study showed that *Scenedemus* sp. and *Chlorella* sp. had a long lag phase in very high concentrations of CO_2 [17]. They further suggested that the dry weight biomass was not affected by variation in the flow rates of air containing elevated CO_2 . This was the result of their use of sea water, which has a strong buffering capacity. In contrary, our experiment use of fresh water containing macro and micro nutrient as artificial growth media, thus the dry weight biomass was affected by variation in the flow rates of air containing elevated CO_2 (Figure 1(b)).

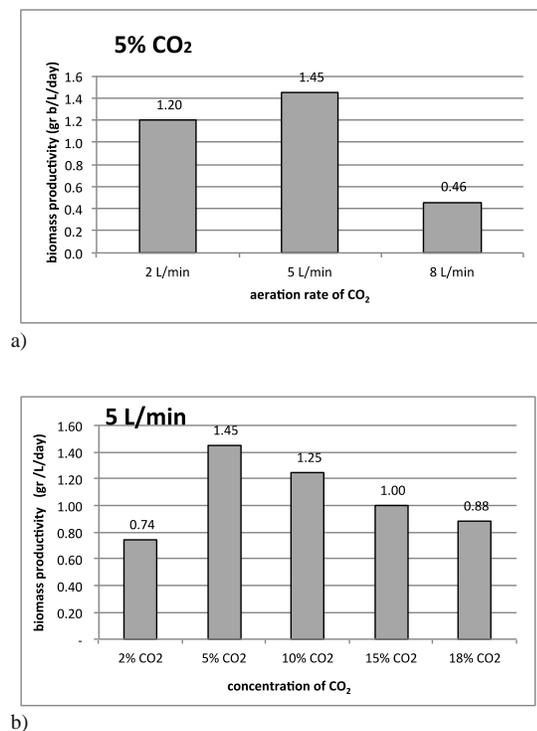
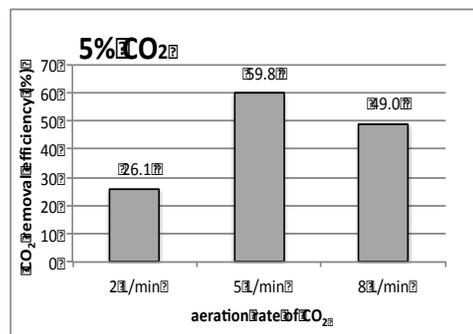


Figure 2 Biomass productivities (a) with variation aeration rate of CO_2 and 5% concentration of CO_2 , (b) with variation concentration of CO_2 (in %; v/v), CO_2 aeration rate of 5 Lmin^{-1} , all were supplied continuously

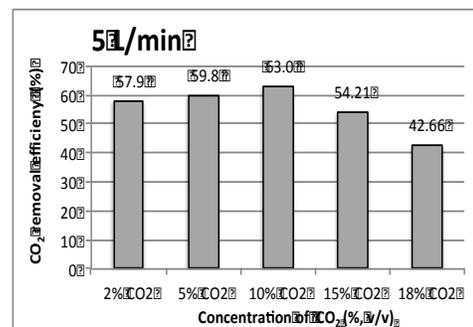
Figure 2(a) shows that biomass productivity in culture that supplied with 5% CO_2 , aeration rate of 2 Lmin^{-1} and 5 Lmin^{-1} were not different significantly. The highest biomass productivity (1.45 $\text{g}\cdot\text{L}^{-1}$), was found at aeration rate of 5 Lmin^{-1} . It means the biomass productivity increased 3-fold highest compare to cultures in aeration rate of 8 Lmin^{-1} . However, Figure 2(b) shows the biomass productivities in culture that supplied more than 5% CO_2 were getting decrease, probably because increasing of CO_2 level become toxicity for growing microalgae.

3.2 Carbon Dioxide Removal Efficiency

CO_2 removal in a vertical bubble column photobioreactor is first marked by a difference in concentration of CO_2 then input into the reactor and the concentration of CO_2 coming out of the reactor. Difference in CO_2 concentration shows that there is a process of removing CO_2 from the air into the microalgae cultivation media (Equation 2).



a)



b)

Figure 3 CO_2 removal efficiency (a) with variation aeration rate of CO_2 and 5% concentration of CO_2 , (b) with variation concentration of CO_2 (in %; v/v), CO_2 aeration rate of 5 Lmin^{-1} , all were supplied continuously

At the same level of CO_2 (5%), although dry weight biomass in aeration rate of 2 Lmin^{-1} was not increasing significantly as compared with 5 Lmin^{-1} , however the CO_2 removal efficiency in aeration rate of 5 Lmin^{-1} was increasing at 2-fold than aeration rate of 2 Lmin^{-1} (Table 1, Figure 3(b)) which shows CO_2 of 8 Lmin^{-1} was lower than at 5 Lmin^{-1} . The CO_2 removal efficiency decreased with increasing gas flow [18].

The reason for this decrease can be explained by the increased gas hold up and turbulence caused in the system because of higher gas flow rates. Under such condition, excessive presence of extremely small gas bubbles were formed which did not participate in the mass transfer of the gas to the liquid phase. Therefore, when the system was highly turbulent there was more gas hold up that forces more CO_2 to leave the photobioreactor system. The highest CO_2 removal efficiency was obtained from the culture which was supplied with CO_2 aeration rate of 5 Lmin^{-1} and 10% CO_2 (Figure 3(b)).

3.3 Carbon Uptake Rate

All algae could take up CO_2 by diffusion, and many had active carbon uptake systems which could take up bicarbonate (HCO_3^-). However, microalgae could not take up the CO_3^{2-} ions [19].

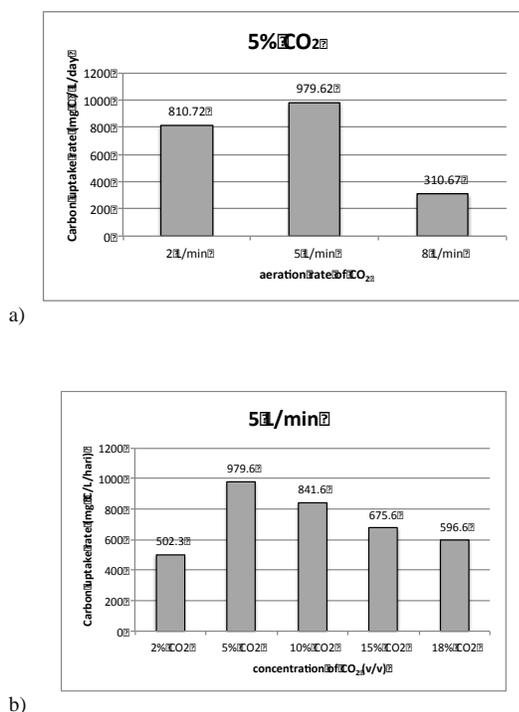


Figure 4 Carbon uptake rate (a) with variation aeration rate of CO₂ and 5% concentration of CO₂, (b) with variation concentration of CO₂ (in %; v/v), CO₂ aeration rate of 5 L/min, all were supplied continuously

Studies have been undertaken to compare low aeration rate (2 Lmin⁻¹) and high aeration rate (8 Lmin⁻¹) with the same high concentration (%; v/v) of CO₂, and it was observed that carbon uptake rate (mg C.L⁻¹.d⁻¹) were higher in low aeration rate than the high aeration rate, which the values were 810.72 and 310.67, respectively (Figure 4(a)). The highest value of carbon uptake rate of 979.62 mg C.L⁻¹.d⁻¹ was recorded from the culture with aeration rate of 5 Lmin⁻¹. In the next experiment (Figure 4(b)) with higher level of CO₂ and the same aeration rate (5 Lmin⁻¹) showed that carbon uptake rate getting decreased in culture that supplied with concentration of CO₂ higher than 5% (v/v).

Many researchers describe that efficiently capturing carbon dioxide and carbon uptake rate from an elevated CO₂ source depends on many factors [23-26], but one of the most limiting factors at present is the ability of the microalgae to capture and fix carbon at a proper concentration to avoid acidification of the medium and crash of the culture, all of which could inhibit the growth of microalgae. It has proven that growth rate (μ) in a culture that was supplied with 10% CO₂ was lower than the culture that was supplied with 5% CO₂, i.e. the values of growth rate were 0.43 and 0.32 respectively (Table 1).

Until the end of the study, the culture supplied with more than 5% CO₂ gave the most unfavorable response compared with 5% and 2% CO₂. High CO₂ concentrations (>5%) generally become toxic to microalgae, presumably because the medium becomes acidic from carbonic acid.

Table 1 Comparison of growth response, carbon dioxide removal efficiency, and carbon uptake rate under variation of carbon dioxide aeration rate

	CO ₂ Aeration rate				Literature
	2Lmin ⁻¹	8Lmin ⁻¹	5Lmin ⁻¹	5Lmin ⁻¹	
CO ₂ removal efficiency (%; v/v)	26.10	49.00	59.80	63.10	▪ 65 [20]; 85 [21] ▪ 45 [22]; 52 [23]
Concentration CO ₂ max that gives highest CO ₂ removal efficiency (%)	5%	5%	5%	10%	▪ 10% [20]; ▪ 10% [21] ▪ 10% [22]; ▪ 10% [23]
Dry weight biomass (g/L)	2.9	3.7	4.9	5.8	▪ 2.046 [23]
Growth rate (μ ; per day)	0.38	0.18	0.43	0.32	▪ ND [20]; ▪ ND [21] ▪ 0.252 [22]; ▪ 0.11 [23]
Biomass productivity (P; g biomass L ⁻¹ .day ⁻¹)	1.20	0.46	1.45	1.25	▪ 0.94 [20]; ▪ 0.632 [21]; ▪ 0.3818 [22] ▪ 0.610 [23]
Carbon consumption rate = Carbon uptake rate (mg Carbon L ⁻¹ .day ⁻¹)	810.72	310.67	979.62	841.6	▪ 1316 [20] ▪ 1367 [21] ▪ 717.8 [22] ▪ 1147 [23]

We assumed that CO₂/O₂-balance is also a prime factor in achieving a higher carbon uptake rate. For this reason, other researchers assume that the carboxyl enzyme, Rubisco, utilizes CO₂ via the Calvin cycle to turn the carbon source into bio-energy, an excess of oxygen may become a problem in the algal culture, not only because it can limit the photosynthesis rate (photorespiration) as well as carbon uptake rate, but also because oxygen radicals may have toxic effects and cause cell membrane damage [27].

■4.0 CONCLUSION

This study showed that aeration rate has a great influence on both removal CO₂ efficiency and carbon uptake rate. These results imply that mix-culture green microalgae can tolerate high concentration of CO₂ at aeration rates of 2 Lmin⁻¹ to 5 Lmin⁻¹. Therefore, when designing CO₂ sequestration systems for microalgae, it should ensure the flow rate is maintained below 5 L.min⁻¹ levels, to allow maximum CO₂ mass transfer into microalgal biomass.

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