ISOLATION OF NANOCELLULOSE FROM JATROPHA WASTE: AN OVERVIEW

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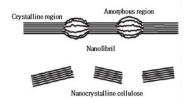
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Graphical abstract



Abstract

Nanocellulose widely used as an additive to improve the quality of composite for medical appliances, electronic and many other applications. The structure can be found in a plant cell wall and established methods are needed for an isolation process. Biomass from plant is commonly selected for this process due to theirs abundance resources. Nanocellulose from jatropha plant will be thoroughly discussed in this paper where several isolation methods will be highlighted.

Keywords: Isolation, jatropha waste, nanocellulose, nanofibrillated, nanocrystalline

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1.0 INTRODUCTION

The high demand on eco-friendly materials lead to many investigation and research on biomaterials. Cellulose is an example of polymer that has been widely research by scientist due to its hydrogen bond network which makes cellulose become a high axial stiffness [1]. The hydrogen bonding also contribute to a stable polymer because of the tendency to form crystalline aggregates. This natural occurring polymer can be found in plant based materials and microorganism.

Anselme Payen in 1838 was first discovered the unique structure of polymer [2]. Since then, many research involving chemical and physical properties of cellulose have been discovered through time. The term 'nanocellulose' is widely used as the result of an isolation process from plant and generally referred as cellulosic nanosize material.

Nanocellulose can be categorized by three types

known in literature as nanofibrillated cellulose, nanocrystalline cellulose and bacterial nanocellulose according to materials and methods used. Nanofibrillated cellulose consists of long and flexible nanosize particles, whereby nanocrystalline cellulose consists of crystal form of nanoparticles. Most of the literatures used microfibrillated cellulose as synonym with nanofibrillated cellulose. However, Moon et al. (2011) described the different between both, where nanofibrillated cellulose is finer in particle diameters than microfibrillated cellulose [3].

Figure 1 shows the difference between nanofibrillated and nanocrystalline celluloses.

The advantages of using nanocellulose have been broadly discussed among the researcher. Samir et al. (2005) listed several advantages of nanocellulose, includes, biodegradable, renewable resources, high specific strength and modulus [4]. Hence, numbers of researches have been done in order to isolate

nanocellulose from various plants. Nanocellulose obtained from plant is more economical than from microorganism, because the latter of nanocellulose

can cause contamination on digestive system [5].

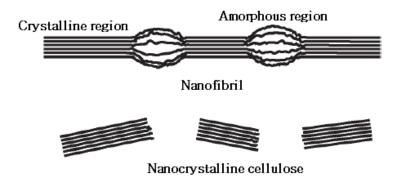


Figure 1 Difference between nanofibril and nanocrystalline cellulose.

Over a decade, isolation of nanocellulose from biomass has gained more attention and interest. Mass production of nanocellulose could be gain since biomass are abundant resources and have high cellulose components. Currently, there is no isolation processes of nanocellulose using jatropha waste have been reported. The characteristics of jatropha waste will be discussed on next subtopic. A

trend and chemical compositions on isolation of nanocellulose from biomass by using various raw materials is shown in Table 1. This paper is an overview on isolation of nanocellulose from jatropha waste that can be used in many fields such as medical and electronic application.

Raw Materials	Year	Chemical Compositions			Reference
		Cellulose (%)	Hemicellulose (%)	Lignin (%)	_
Banana pseudeostem	2008	64.04 <u>+</u> 2.8	18.60 <u>+</u> 1.6	4.90 <u>+</u> 0.7	[6]
Mulberry Barks	2009	37.4 <u>+</u> 2.3	25.3 <u>+</u> 2.5	10.0 <u>+</u> 0.8	[7]
Pineapple Leaf	2010	81.3 <u>+</u> 2.5	12.3 <u>+</u> 1.4	3.5 <u>+</u> 0.6	[8]
Corncob	2013	31.2 <u>+</u> 3.1	43.1 <u>+</u> 4.0	16.5 <u>+</u> 2.0	[9]
Oil Palm Empty Fruit Bunch	2014	45.0 <u>+</u> 5.0	25.0 <u>+</u> 5.0	25.0 <u>+</u> 5.0	[10]

Table 1 A trend and chemical compositions of nanocellulose from biomass.

2.0 JATROPHA WASTE AS AN ABUNDANT SOURCE OF CELLULOSE

Jatropha (Jatropha curcas L.) or common name, physic nut, has been known as multipurpose plant. This plant belongs to Euphorbiaceae family and can be found in South America, Africa, India, and South Asia. Jatropha is a drought resistant and can cultivate in almost any type of soils, therefore, this plant is easily established.

Several researchers have been focused on jatropha as second generation fuel. As palm oil, jatropha can produce oil which is important for renewable energy sources. In addition, palm oil is an edible which will be used for food and cooking

based material. However, jatropha is non edible as palm oil, thus very useful as non food feedstock for oil extraction.

As a woody plant, biomass from jatropha consists of lignocelluloses, a combination of cellulose, hemicelluloses and lignin. Previous research has been done to determine the cellulose percentage in different part of jatropha plant. Based on Table 2, jatropha waste has the highest percentage of cellulose compared to other biomass. Eventhough the percentage shown was 56% but by using chemical treatment, this percentage can be increased.

This percentage is higher as compared to oil palm empty fruit bunch in Table 1. The compositions of

cellulose indicate the potential that nanocellulose can be isolated from jatropha waste. Recent studies reported that, the jatropha waste can be utilized from oil extraction using jatropha seeds. There are many types of waste that generate from biodiesel production, including husk, shell and seed cake.

Type of fibre	Cellullose (%)	Lignin (%)	Hemicellulose (%)	Reference
Jatropha Waste	56.31	23.91	17.47	[11]
Hull	42.8 <u>+</u> 0.6	9.6 <u>+</u> 0.6	14.7 <u>+</u> 0.5	[12]
Wood Component of Shell	22.29	47.60	23.84	[13]
Stem	42.99	24.11	19.11	[14]
Shell	33.75	11.90	9.70	[15]
Fruit Coat	13.11	28.91	7.69	[16]

Table 2 The composition of difference part of jatropha plant

3.0 NANOCELLULOSE RECOVERY FROM PLANT WASTE

There are two types of nanocellulose that can be formed from plant, which are nanofibrillated cellulose or nanocrystalline cellulose. The recovery yield depends on the method used. Several methods have been discussed by researcher to obtain nanocellulose. Several novelty methods have been found in order to achieve and isolates nanocellulose from plant. However, each method has its own advantages and disadvantage compared to others. The method used depends on the level of difficulty to isolate the cellulose from the plant. High concentration of chemical reagent will be required if the lignin content in the plant is higher than usual [17].

Generally, there are two stages of producing plant nanocellulose from source pretreatment and chemical or mechanical methods [18]. Nanofibrillated cellulose is generally isolated by using mechanical method, meanwhile, nanocrystalline cellulose are generated from chemical method. The most common method use to produce nanofibrillated cellulose is by using highpressure homogenizer. This method was applied by Herrick et al. (1983) and Turbak et al. (1983), where they first discovered the nanofibrillated cellulose [19, 20]. This method involved homogenizing cellulose into slurry that is usually suspended in water, by using high pressure applied to the nozzle of the homogenizer. Cryocrushing is an alternative mechanical method to produce nanocellulose. This method used liquid nitrogen to freeze the sample and high impact is applied to crush the sample through mortar and pestle. Alemdar and Sain (2008), applied this technique prior of using laboratory defibrillator [21]. They found that by applying this technique, the size of nanofibrillated cellulose can be reduced.

As for nanocrystalline cellulose, a pretreatment is important to isolate cellulose from hemicelluloses and other impurities component. After a pretreatment process, acid hydrolysis is used to remove amorphous region in order to obtain high crystallite particle or nanocrystalline cellulose. Acid hydrolysis involve high concentration of acid such as hydrochloric acid (HCI) and sulphuric acid (H₂SO₄) as a medium. During this process, only crystalline region will remain because of their resistance against acid [22]. Steam explosion is a technique that can also be used to isolate cellulose from plant fibres. This technique uses hot steam and pressure, followed by a fast decompression to derive cellulose nanofibrils from plants. Cherian et al. (2008), had use steam explosion in alkaline medium, and subsequently acidic medium to derive nanocellulose from banana fibers [6]. This method is an efficacious method to isolate cellulose nanofibrils from biomass due to less hazardous of chemical used and low environmental impact [6].

Several steps to isolate nanocrystalline cellulose from biomass were described by Brinchi et al. (2013), starting from pretreatment, followed by acid hydrolysis, dialysis and sonication [18]. Sonication plays an important role to stabilize the dispersion of nanocrystals. Combination of those methods resulted on more promising cellulose in term of length and diameters. Previous researches on using biomass to produce nanocellulose are shown in Table 1.

Width Type of Length **Authors** Methods nanocellulose (nm) (nm) Steam explosion in alkaline medium and Nanocrystalline Cherian et al. (2010) [8] 200 - 3005 - 60bleaching, followed Cellulose by acidic medium Alkali treatment, acid hydrolysis and bleaching, refining Nanofibrillated Wang & Sain (2007) [23] Several µm 50 - 100Cellulose and beating, followed by high pressure defibrillation Alkali and bleaching treatment, acid Nanofibrillated Pelissari et al. (2014) [24] 375.2 - 454.9 10.9 - 22.6hydrolysis with high-Cellulose pressure homogenization Alkali and bleaching treatment, blending, Nanocrystalline Silvério et al. (2013) [9] 166.6 - 2553.07 - 5.23

Cellulose

Cellulose

Cellulose

Nanocrystalline

Nanocrystalline

Table 3 Previous research on using biomass for nanocellulose

4.0 CONCLUSION

Li et al. (2009) [7]

Rosa et al. (2010) [25]

An overview shows the potential of using jatropha waste as raw materials for isolation of nanocellulose based on cellulose percentage that is above than 50% which will require less concentration of chemical reagent. The type of nanocellulose produce depends on the method used, whereby a combination method of chemical and mechanical will result with shorter nanocellulose in term of length and diameter.

acid hydrolysis and

Alkali and bleaching

Alkali and bleaching treatment, acid

treatment, acid

hydrolysis and

sonication

sonication

hydrolysis

Acknowledgement

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References

- [1] Cherian, B. M., Leão, A. L., De Souza, S. F., Costa, L. M. M., De Olyveira, G. M., Kottaisamy, M., Nagarajan, E. R. and Thomas, S. 2011. Cellulose Nanocomposites With Nanofibres Isolated From Pineapple Leaf Fibers For Medical Applications. Carbohydr. Polym. 86(4): 1790–1798.
- [2] Payen, A. 1838. Mémoire Sur La Composition Du Tissu Propre Des Plantes Et Du Ligneux. Comptes Rendus. 7: 1052–1056

[3] Moon, R., Martini, A. and Nairn, J. 2011. Cellulose Nanomaterials Review: Structure, Properties And Nanocomposites. Soc. Rev. 40(7): 3941–3994.

4 - 8.3

20 - 40

58 - 515

400 - 500

- [4] Samir, M. A., Alloin, F. and Dufresne, A. 2005. Review Of Recent Research Into Cellulosic Whiskers, Their Properties And Their Application In Nanocomposite Field. Biomacromolecules. 612–626.
- [5] Bhatnagar, A. and Sain, M. 2005. Processing of Cellulose Nanofiber-reinforced Composites. J. Reinf. Plast. Compos. 24(12): 1259–1268.
- [6] Cherian, B. M., Pothan, L. A., Nguyen-Chung, T., Mennig, G., Kottaisamy, M. and Thomas, S. 2008. A Novel Method For The Synthesis Of Cellulose Nanofibril Whiskers From Banana Fibers And Characterization. Journal Of Agricultural And Food Chemistry. 56(14): 5617–5627.
- [7] Li, R., Fei, J., Cai, Y., Li, Y., Feng, J. and Yao, J. 2009. Cellulose Whiskers Extracted From Mulberry: A Novel Biomass Production. Carbohydr. Polym. 76(1): 94–99.
- [8] Cherian, B. M., Leão, A. L., de Souza, S. F., Thomas, S., Pothan, L. A. and Kottaisamy, M. 2010. Isolation Of Nanocellulose From Pineapple Leaf Fibres By Steam Explosion. Carbohydr. Polym. 81(3): 720–725.
- [9] Silvério, H. A., Flauzino Neto, W. P., Dantas, N. O. and Pasquini, D. 2013. Extraction And Characterization Of Cellulose Nanocrystals From Corncob For Application As Reinforcing Agent In Nanocomposites. *Ind. Crops Prod.* 44: 427–436.
- [10] Lani, N. S., Ngadi, N., Johari, a. and Jusoh, M. 2014. Isolation , Characterization , and Application of Nanocellulose from Oil Palm Empty Fruit Bunch Fiber as Nanocomposites, J. Nanomater. 1–9.
- [11] Sricharoenchaikul, V. Marukatat, C. and Atong, D. 2007. Fuel Production From Physic Nut (Jatropha Curcas L.) Waste By Fixed-Bed Pyrolysis Process. Thail. J. 1–6..
- [12] Jiang, L.-Q., Fang, Z., Guo, F. and Yang, L. 2012. Production Of 2,3-Butanediol From Acid Hydrolysates Of Jatropha Hulls With Klebsiella Oxytoca. *Bioresour. Technol.* 107: 405–410.

- [13] Wever, D.-A. Z., Heeres, H. J. and Broekhuis, A. A. 2012. Characterization Of Physic Nut (Jatropha Curcas L.) Shells. Biomass and Bioenergy, 37: 177–187.
- [14] Vaithanomsat, P. and Apiwatanapiwat, W. 2009. Feasibility Study On Vanillin Production From Jatropha Curcas Stem Using Steam Explosion As A Pretreatment. Inter J Chem Biolo Engr. 839–842.
- [15] Singh, R., Vyas, D., Srivastava, N. and Narra, M. 2008. SPRERI Experience On Holistic Approach To Utilize All Parts Of Jatropha Curcas Fruit For Energy. Renew. Energy. 33: 1868–1873.
- [16] Dhanya, M., Gupta, N. and Joshi, H. 2009. Biogas Potentiality Of Agro-Wastes Jatropha Fruit Coat. Proc. World Acad. Sci. 432–436.
- [17] Nascimento, D. M., Almeida, J. S., Dias, A. F., Figueirêdo, M. C. B., Morais, J. P. S., Feitosa, J. P. a. and de F Rosa, M. 2014. A Novel Green Approach For The Preparation Of Cellulose Nanowhiskers From White Coir. Carbohydr. Polym. 110: 456–463.
- [18] Brinchi, L., Cotana, F., Fortunati, E. and Kenny, J. M. 2013. Production Of Nanocrystalline Cellulose From Lignocellulosic Biomass: Technology And Applications. Carbohydrate Polymers. 94(1): 154–169.
- [19] Herrick, F. W., Casebier, R. L. K., Hamilton, J. and Sandberg, K. R. 1983. Microfibrillated Cellulose: Morphology and

- Accessibility. J. Appl. Polym. Sci. Appl. Polym. Symp. 37: 797–813.
- [20] Turbak, A. F., Snyder, F. W. and Sandberg, K. R. 1983. Microfibrillated Cellulose, A New Cellulose Product: Properties, Uses, and Commercial Potential. J. Appl. Polym. Sci. Appl. Polym. Symp. 37: 815–827.
- [21] Alemdar, A. and Sain, M. 2008. Isolation And Characterization Of Nanofibers From Agricultural Residues-Wheat Straw And Soy Hulls. *Bioresour. Technol.* 99: 1664–1671.
- [22] Habibi, Y. Lucia, L. a. and Rojas, O. J. 2010. Cellulose Nanocrystals: Chemistry, Self-Assembly, And Applications. Chem. Rev. 110(6): 3479–3500.
- [23] Wang, B. and Sain, M. 2007. Isolation Of Nanofibers From Soybean Source And Their Reinforcing Capability On Synthetic Polymers. 67: 2521–2527.
- [24] Pelissari, F., do P., Sobral, A. and Menegalli, F. 2014. Isolation And Characterization Of Cellulose Nanofibers From Banana Peels. Cellulose. 417–432.
- [25] Rosa, M. F., Medeiros, E. S., Malmonge, J. a., Gregorski, K. S., Wood, D. F., Mattoso, L. H. C., Glenn, G., Orts, W. J. and Imam, S. H. 2010. Cellulose Nanowhiskers From Coconut Husk Fibers: Effect Of Preparation Conditions On Their Thermal And Morphological Behavior. Carbohydr. Polym. 81(1): 83–92.