# Jurnal Teknologi

# EFFECTS OF Justicia gendarussa ETHANOLIC EXTRACT ON OSTEOBLASTIC ACTIVITY OF MC3T3-E1 CELL

Kavita Supparmaniam, Siti Pauliena Mohd Bohari\*

Department of Biotechnology and Medical Engineering, Faculty of Bioscience and Medical Engineering, Universiti Teknologi Malaysia, Johor, Malaysia Article history
Received
23 March 2015
Received in revised form
14 July 2015
Accepted
21 July 2015

\*Corresponding author pauliena@fbb.utm.my

# **Graphical abstract**



# Abstract

Justicia gendarussa (Acanthaceae) or commonly known as Gendarussa has traditionally been used to treat bone fractures. Bone fracture is a clinical condition that need bone repair and new bone formation. To date, the mechanism of Justicia gendarussa acting in enhancing the bone mineralization has not been proven scientifically. The present study aimed to investigate the cytotoxicity and alkaline phosphatase (ALP) activity on osteoblast cells when treated with Justicia gendarussa ethanolic leaves extract. MTT and ALP assays were performed on osteoblast cells after being treated with different concentrations of the extract. For cell viability, the result showed that  $IC_{50}$  value of the osteoblast cells was 89.1 $\mu$ g/ml. While, ALP assay is used as a biochemical marker for early detection of osteoblast mineralization. The highest amount of ALP activity was at the 37.5  $\mu$ g/ml when compared to the control. From this study, it shows that Justicia gendarussa has potential in enhancing bone mineralization during the bone repair process.

Keywords: Bone mineralization, Justicia gendarussa, osteoblast, MTT assay, ALP assay

# **Abstrak**

Justicia gendarussa (Acanthaceae) atau lebih dikenali sebagai Gendarussa telah digunakan secara tradisional untuk merawat tulang patah. Tulang patah adalah keadaan klinikal yang memerlukan penyembuhan dan pembentukan tulang baru. Sehingga kini, mekanisma bagi Justicia gendarussa meningkatkan pemineralan tulang masih belum dibuktikan secara saintifik. Kajian ini bertujuan untuk mengkaji kesan sitotoksik (MTT) dan aktiviti 'alkaline phosphatase' (ALP) terhadap sel-sel osteoblast setelah dirawat dengan ekstrak ethanol yang diperolehi dari daun Justicia gendarussa. Asai MTT dan ALP telah dijalankan dengan mengunakan sel osteoblast selepas dirawat dengan ekstrak yang berbeza kepekatannya. Asai MTT digunakan untuk kemandirian sel hidup dan hasilnya menunjukkan bahawa nilai IC<sub>50</sub> untuk sel-sel osteoblast adalah 89.1 μg/ml. Manakala, asai ALP digunakan sebagai penanda biokimia untuk pengesanan awal pemineralan osteoblast. Jumlah tertinggi aktiviti ALP adalah pada 37.5 μg/ml dan berbanding dengan sel osteoblast yang tidak dirawat dengan esktrak, aktiviti ALP tinggi pada kepekatan ini. Kesimpulannya, Justicia gendarussa berpotensi dalam meningkatkan pemineralan tulang semasa proses pemulihan.

Kata kunci: Pemineralan tulang, Justicia gendarussa, osteoblast, MTT asai, ALP asai

© 2015 Penerbit UTM Press. All rights reserved

#### 1.0 INTRODUCTION

Justicia gendarussa is belongs to the family of Acanthaceae and commonly known as Gendarussa [1]. It is found in tropical and subtropical throughout Asian countries like India, Malaysia, Indonesia and Sri Lanka [2]. Phytochemical research on the leaves of this plant revealed that alkaloids, amino acids, aromatic amines, flavonoids, triterpinoidal, saponins [1] tannins, justicin and steroids are present in this plant [3]. This plant has been used traditionally to treat chronic rheumatism, inflammations, bronchitis, vaginal discharges, dyspepsia, eye diseases, muscle pain, lumbago, headache, earache, hemiplegia, hair growth promotion, leucoderma, asthma, antiseptic, haemostatic, nasal bleeding, injuries and fever [3-10]. Previous researches have shown that this plant has antimicrobial activity, antihelminth activity, larvacidal and adulticidal activities, in vitro HIV type 1 reverse transcriptase inhibitory activity, antisickling activity on sickle cell, antinociceptive activity, antianaioaenic effect, anti-inflammatory analgesic activity, antioxidant and hepatoprotective potential and anti-arthritic potential [3,11-21]. However, J. gendarussa only has been used traditionally to cure bone fracture [6].

Bone is the body framework and also a structural support for an individual. Bone is able to regenerate and form new osseous tissue whenever the resorption process occur in the bone. Bone fracture is one of the clinical condition that caused either by trauma like accidents and few diseases like osteoporosis, tumor and deficiency of calcium or vitamin D [22]. It can occurs in any type of bones in the body and may cause swelling, protruding bone or blood under skin, numbness, extreme pain, tenderness and paralysis. Basically, there are two types of bone fracture which are hairline or simple fracture and compound fracture. Simple fracture occurs when the skin is still intact in the fractured area while, compound fracture occurs when the skin breaks open due to the broken bones [22]. The treatment for bone fracture can be done by surgical like implant fixation and nonsurgical like usage of drugs [23]. Although there are number of treatment, the healing of bone fracture usually takes a longer period, ranging from six to eight weeks [24].

Traditional herbal medicines which have been used in medical practice might play important role in bone fracture healing [25]. To our knowledge, there is no experimental evidence to support on the mechanism of *J. gendarussa* acting on osteoblast cells for detecting the mineralization process. Thus, the aim of this study is to investigate the cytotoxicity and alkaline phosphatase (ALP) activity of osteoblast cells when treated with ethanolic leaves extract of *J. gendarussa*.

## 2.0 EXPERIMENTAL

#### 2.1 Plant Materials

J.gendarussa plants were bought from Nursery Pak Ali in Skudai, Johor, Malaysia. These plants were raised in pots containing soil under greenhouse conditions at Faculty of Biosciences and Medical Engineering, UTM.

#### 2.2 Extraction

The young and green leaves of *J.gendarussa* plants were collected and cleaned thoroughly under running tap water. The leaves were cut into small pieces, air dried and powdered. The powdered leaves were macerated with 96% ethanol for three days at room temperature [26]. The liquid extract was filtered using Advantec filter paper (ADVANTEC Toyo Roshi International, Inc., Dublin, CA, USA) and evaporated under reduced pressure using a rotary evaporator (EYELA N-1110, EYELA, Tokyo, Japan) and the procedure above was repeated few times. The sample yields a gummy ethanol extract. The sample was dried in freeze dryer (Beta 2-4 LD plus LT, Martin Christ, Germany) until complete dryness and yield powder form [27].

#### 2.3 Materials

Minimum Essential Media alpha medium (a-MEM), fetal bovine serum (FBS), trypsin and penicillin/streptomycin were purchased from Gibco Company (Bio-Diagnostic Sdn Bhd, Petaling Jaya, Selangor, Malaysia), 3-(4,5-dimethylthiazol-2,5-diphenyl tetrazolium bromide (MTT) from Sigma-Aldrich Group (Medigene Sdn Bhd, Puchong, Selangor, Malaysia) and ethanol (96%, Qrec, Malaysia)

# 2.4 Cell Cultures

The mouse osteoblast cell line (MC3T3-E1) was purchased from American Type Culture Collection (ATCC). MC3T3-E1 cells were cultured in alpha minimal essential medium (a-MEM) containing 10% fetal bovine serum and 1% penicillin/streptomycin in a humidified incubator with mixture of 95% air and 5%  $\rm CO_2$  at 37°C. The medium was changed every 3 to 5 days prior to use [28].

# 2.5 MTT Assay

Cell optimization was done using different cell densities  $(2\times10^3-1\times10^5)$ . Based on the result,  $1\times10^4$  cells/ml seeding density were selected for MC3T3-E1 cell and seeded in 96-well plate [29]. After 24 hours of incubation, the cells were treated with different concentrations of *J. gendarussa* ethanolic leaves extract  $(7.81-1000 \ \mu\text{g/mL})$  and incubated for 72 hours. MTT (3-(4,5-dimethylthiazol-2,5-diphenyl)

tetrazolium bromide) was dissolved in phosphate buffer saline at 5 mg/mL. 20  $\mu L$  of the solution was added to each well and the plate was incubated at 37°C for 4 hours. Then, medium was discarded and 225  $\mu L$  of 1N HCl/isopropanol buffer was added to each wells to dissolve the purple crystals [1]. The plate was read at the wavelength of 560 nm using Promega GloMax microplate reader (Promega, Wisconsin, USA) [30]. The results were recorded as percentage of cell viability using the formula given as below [31] .

% Cell viability =  $(Abs ext{ of test} - Abs ext{ of blank})$   $\times 100$  (Abs of control – Abs of blank

Where, Abs = Absorbance

# 2.6 Alkaline Phosphatase (ALP) Assay

Cell optimization was done using different cell densities  $(4\times10^4-1\times10^5)$ . From the result,  $4\times10^4$  cells/mL seeding density were chosen for MC3T3-E1 cell and seeded in 24-well plate. After 24 hours of incubation, cells were treated with different concentrations of J. gendarussa ethanolic extract (9.38-15 µg/mL) for 72 hours. Then, the cells were washed once with phosphate buffer saline and lysed in 0.5 mL of 0.5% Triton X-100 in each well for 30 minutes at room temperature. The lysate were centrifuged at 3000 rpm for 10 minutes and the supernatant were used for the determination of ALP. ALP activity was assayed with a QuantiChrom<sup>TM</sup> Alkaline Phosphatase Assay Kit (DALP-250; BioAssay System, CA) according to manufacturer's protocol. Optical density was determined at 405 nm by using a BioTek microplate reader (BioTek Instruments, Winooski, VT, USA) [32].

# 2.7 Statistical Analysis

The results expressed as a mean±standard deviation (SD) from triplicate values. SPSS16.0 software (SPSS 16.0 for Windows Evaluation Version software, SPSS Inc., USA) was used to analyze the data. The normality of the data was determined using the Shapiro-Wilk test. The statistical significance was evaluated using an independent *t*-test for normal data while the Mann - Whitney test was used for nonnormal data. A *p*-value less than 0.05 were considered significant [1].

### 3.0 RESULTS AND DISCUSSION

In this study, cytotoxicity and ALP activity of *J.gendarussa* ethanolic leaves extract were evaluated using MTT and ALP assay. MTT assay is the colorimetric method in which yellow MTT dye was reduced by succinic dehydrogenase in the mitochondria of viable cells to form purple formazan crystal [33].

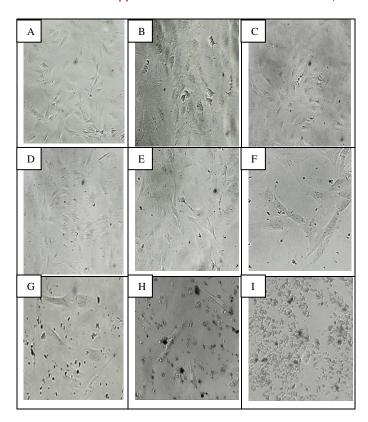
Based on Table 1, the percentage of viability significantly decreases with the increasing of J. gendarussa ethanolic leaves extract concentration. The result indicates that the cytotoxic effect increases with the J. gendarussa ethanolic extract concentration. IC50 value is a 50 % reduction in viability of cells [1] and it is determined by plotting a line graph of percentage of viability against Log (concentration of extract) [31]. Using the graph, the Log value of the 50 % of cell viability is determined and antilog of that value gives the exact IC<sub>50</sub> value [31]. The IC<sub>50</sub> value of J. gendarussa towards MC3T3-E1 cells is 89.1 µg/ml. Previous study stated that, IC50 value below than 20 µg/ml is toxic to the cell, ranging between 21-40 µg/ml is less toxic and not toxic if the  $IC_{50}$  value is above than 41 µg/ml [1]. The ethanolic extract of J.gendarussa is considered not toxic to the osteoblast cell since the IC $_{50}$  value is above 41  $\mu g/ml$ .

**Table 1** Percentage of cell viability of MC3T3-E1 cells treated with different concentrations of *J gendarussa* ethanolic leaves extract

Concentration (µg/ml)	Control	7.81	15.63	31.25	62.5	125	250	500	1000
		***	**	***	***	*	***	***	*
% of cell viability	100	72.71	70.94	64.99	61.29	40.96	26.34	16.02	8.68
	±0.008	±0.001	±0.009	±0.009	±0.001	±0.005	±0.009	±0.318	±0.012
IC₅o(µg/ml)					89.1				

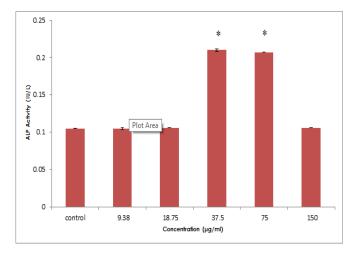
Values are mean  $\pm$  STDEV for three replicates; \*p<0.05, \*\*p<0.01 and \*\*\*p<0.001 compared with control

Figure 1 shows the morphology and confluency of the MC 3T3-E1 cells in different concentration of J. gendarussa ethanolic extract. Based on the figure, the confluency of the cells decreases when the concentration of the extract increases. This indicates a concentration dependent manner of J.gendarussa ethanolic extract towards osteoblasts. Thus, the cells are able to arow and maintain their morphology at lower concentration (500  $\mu$ g/mL - 7.81  $\mu$ g/mL). At the highest concentration (1000 µg/mL), morphology of the MC3T3-E1 cells changed into a rounded shape when compared to the control.



**Figure 1** Morphology of MC 3T3-E1 cells after 72 hours treated with different concentration J. gendarussa ethanolic extract (200× magnification). (A) control, without any treatment; (B) J. gendarussa at 7.81 μg/mL; (C) J. gendarussa at 15.63 μg/mL; (D) J. gendarussa at 31.25 μg/mL; (E) J. gendarussa at 62.5 μg/mL; (F) J. gendarussa at 125 μg/mL; (H) J. gendarussa at 500 μg/mL; (I) J. gendarussa at 1000 μg/mLl.

The effect of J. gendarussa ethanolic extract on ALP activity was expressed in Figure 2. The result showed that ALP activity is significantly higher than control at the concentration of 37.5  $\mu$ g/mL and 75 ug/mL. This suggests that at lower concentration (below than 100 µg/mL), this plant extract could stimulate the ALP activity in osteoblast cell. While, J. gendarussa ethanolic extract treated group at the concentration of 9.38  $\mu$ g/mL, 18.75  $\mu$ g/mL and 150 µg/mL showed not much difference in ALP activity when compared with control. This shows that at these concentrations there is no stimulation of ALP activity. The J. gendarussa ethanolic extract at the concentration of 150  $\mu$ g/mL is above the IC<sub>50</sub> value 89.1 µg/mL which that concentration has more than 50% of inhibited cells growth. The concentration of 150 µg/mL is chosen in this experiment to show that higher ALP is not detected when MC3T3-E1 cells growth are inhibited. As ALP is an early marker which used to detect osteoblast cell differentiation and the ALP activity is elevated when there is increased in osteoblast cell differentiation [34-35]. Differentiation of osteoblast cell to become osteocyte is the final phase of differention, where the osteocyte cells embedded in the mineralized bone matrix and forms bone [36]. From this, we can suggest that J. gendarussa can increase osteoblastic differentiation into osteocyte at the a specific concentration. Since early stage is a necessary step in bone mineralization, the enhancing effect of J. gendarussa may stimulate the bone fracture healing.



**Figure 2** ALP activity of osteoblast cell line treated with different concentrations of J. gendarussa ethanolic leaves extract. Values are expressed as the mean  $\pm$  SD for three replicates, \*p<0.05 compared with control

# 4.0 CONCLUSION

As a conclusion, this study showed that J.gendarussa has potential in increasing the ALP activity in osteoblast cells. This suggested that J. gendarussa treatment can stimulate early stage of mineralization and thus, helps osteoblast cell differentiation. Therefore, J. gendarussa treatment may be favorable for the bone fracture healing, with a potential mechanism of stimulating the ALP activity in osteoblast cell. Further studies are needed to explore the exact mechanism of this plant acting on the osteoblast cells during the bone healing process.

# Acknowledgement

The authors wish to thank Ministry of Higher Education (MOHE) for their funding under Research University Grant (VOT 4F344) and MyBrain 15 scholarship. We also thank Faculty of Biosciences and Medical Engineering, Universiti Teknologi Malaysia (UTM) for their facilities and services provided.

#### References

[1] Ayob, Z., Samad, A. A., and Bohari, S. P. M. 2013. Cytotoxicity Activities in Local Justicia gendarussa Crude Extracts against Human Cancer Cell Lines. Jurnal Teknologi. 64(2): 45-52.

- [2] Janarthanam, B., and Sumathi, E. 2010. In Vitro Regeneration of Justicia gendarussa Burm. f. Libyan Agriculture Research Center Journal Internation. 1(5): 284-287.
- [3] Sugumaran, P., Kowsalya, N., Karthic, R., and Seshadri, S. 2013. Biomass Production And Antibacterial Activity of Justicia gendarussa Burm. f.—A Valuable Medicinal Plant. Journal of Tropical Life Science. 3(1): 8-13.
- [4] Ahmad, F. B., and Holdsworth, D. K. 2003. Medicinal plants of Sabah, East Malaysia-Part I. *Pharmaceutical Biology*. 41(5): 340-346.
- [5] Anonymous. 1959. The Wealth of India. CSIR Publications. New Dehli. ISBN: 81-85038-00-7, 312.
- [6] Das, A. K., Dutta, B., and Sharma, G. 2008. Medicinal Plants Used by Different Tribes of Cachar District, Assam. Indian Journal of Traditional Knowledge. 7(3): 446-454.
- [7] Dolui, A., Sharma, H., Marein, T. B., & Lalhriatpuii, T. 2004. Folk Herbal Remedies from Meghalaya. *Indian Journal of Traditional Knowledge*. 3(4): 358-364.
- [8] Madhu, V., and Naik, D. 2009. Ethnomedicinal Uses of Leaf Preparations in Adilabad District, Andhra Pradesh, India. Ethnobotanical Leaflets. 2009(11): 1.
- [9] Rahman, A., Alam, M., Ahmad, S., Naderuzzaman, A., and Islam, A. 2012. An Ethnobotanical Portrait of a Village: Koikuri, Dinajpur with Reference to Medicinal Plants. International Journal of Biosciences. 2(7): 1-10.
- [10] Zheng, X.-I., and Xing, F.-w. 2009. Ethnobotanical Study on Medicinal Plants Around Mt. Yinggeling, Hainan Island, China. Journal of Ethnopharmacology. 124(2): 197-210.
- [11] Sudhanandh, V., Arjun, J., Faisal, A., Ani, M., Renjini, V., and Babu, K. N. 2012. In-Vitro Anti Bacterial Screening of Selected Folklore Indian Medicinal Plants with Few Clinical Pathogens. Indian Journal of Pharmaceutical Education and Research. 46(2): 174-178.
- [12] Sharma, K., Saikia, R., Kotoky, J., Kalita, J., and Devi, R. 2011. Antifungal Activity of Solanum Melongena L, Lawsonia Inermis L. and Justicia gendarussa B. against Dermatophytes. International Journal of PharmTech Research. 3(3):1635-1640.
- [13] Saha, M. R., Debnath, P. C., Rahman, M. A., and Islam, M. A. U. 2012. Evaluation of in Vitro Anthelmintic Activities of Leaf and Stem Extracts of Justicia gendarussa. Bangladesh Journal of Pharmacology. 7(1): 50-53.
- [14] Senthilkumar, N., Varma, P., and Gurusubramanian, G. 2009. Larvicidal and Adulticidal Activities of Some Medicinal Plants Against the Malarial Vector, Anopheles stephensi (Liston). Parasitology Research. 104(2): 237-244.
- [15] Woradulayapinij, W., Soonthornchareonnon, N., and Wiwat, C. 2005. In Vitro HIV Type 1 Reverse Transcriptase Inhibitory Activities of Thai Medicinal Plants and Canna indica L. rhizomes. Journal of Ethnopharmacology. 101(1): 84-89.
- [16] Mpiana, P. 2010. Antisickling Activity of Three Species of Justicia from Kisangani (DR Congo): J. tenella,/i>, J. gendarussa and J. insularis. International Journal of Biological and Chemical Sciences. 4(6): 1953-1961.
- [17] Ratnasooriya, W., Deraniyagala, S., and Dehigaspitiya, D. 2007. Antinociceptive Activity and Toxicological Study of Aqueous Leaf Extract of Justicia gendarussa Burm. F. in rats. Pharmacognosy Magazine. 3(11):145-155.
- [18] Periyanayagam, K., Umamaheswari, B., Suseela, L., Padmini, M., and Ismail, M. 2009. Evaluation of Antiangiogenic Effect of the Leaves of Justicia gendarussa Burm. (Acanthaceae) by Chrio Allontoic

- Membrane Method. American Journal of Infectious Disease. 5(3): 180-182.
- [19] Jothimanivannan, C., Kumar, R., and Subramanian, N. 2010. Anti-inflammatory and Analgesic Activities of Ethanol Extract of Aerial Parts of Justicia gendarussa Burm. International Journal of Pharmacology. 6(3): 278-283
- [20] Krishna, K., Mruthunjaya, K., and Patel, J. A. 2010. Antioxidant and Hepatoprotective Potential of Stem Methanolic Extract of Justicia gendarussa Burm. International Journal of Pharmacology. 6(2): 72-80.
- [21] Paval, J., Kaitheri, S. K., Potu, B. K., Govindan, S., Kumar, R. S., Narayanan, S. N., and Moorkoth, S. 2009. Anti-arthritic Potential of the Plant Justicia gendarussa Burm F. Clinics. 64(4): 357-362.
- [22] Singla, C., Drabu, S., Verma, R., Dhiman, A., and Sharma, A. 2011. Recent Update on Proficient Bone Fracture Revivifying Herbs. International Research Journal of Pharmacy. 2(11): 3-5.
- [23] Brandi, M. L. 2012. Drugs for Bone Healing. Expert Opinion on Investigational Drugs. 21(8): 1169-1176.
- [24] Singh, V., Singh, N., Pal, U., Dhasmana, S., Mohammad, S., and Singh, N. 2011. Clinical Evaluation of Cissus Quadrangularis and Moringa Oleifera and Osteoseal as Osteogenic Agents in Mandibular Fracture. National Journal of Maxillofacial Surgery. 2(2): 132-132.
- [25] Weyermann, J., Lochmann, D., and Zimmer, A. 2005. A Practical Note on the Use of Cytotoxicity Assays. International Journal of Pharmaceutics. 288(2): 369-376
- [26] Shikha, P., Latha, P., Suja, S., Anuja, G., Shyamal, S., Shine, V., Rajasekharan, S. 2010. Anti-inflammatory and Antinociceptive Activity of Justicia gendarussa Burm. f. leaves. Indian Journal of Natural Products and Resources. 1(4): 456-461.
- [27] Basah, K., Elya, B., Amin, J., and Julian, M. I. 2011. Activity of Ethanolic Extract from Justicia gendarussa burm. Leaves on Decreasing the Uric Acid Plasma. MAKARA of Science Series. 15(1): 67-70.
- [28] Shona Pek, Y., & Wan, A. C. 2011. Characterization of Early Extracellular Matrix Secretions in a Three-Dimensional Thixotropic Cell Culture System. Acta biomaterialia. 7(11): 3981-3987.
- [29] Campos, D. M., Anselme, K., & Soares, G. D. d. A. 2012. In Vitro Biological Evaluation of 3-D Hydroxyapatite/Collagen (50/50 wt.(%)) Scaffolds. Materials Research. 15(1): 151-158.
- [30] Ait-Mohamed, O., Battisti, V., Joliot, V., Fritsch, L., Pontis, J., Medjkane, S., Rholam, M. 2011. Acetonic Extract of Buxus Sempervirens Induces Cell Cycle Arrest, Apoptosis and Autophagy in Breast Cancer Cells. PloS One. 6(9): e24537.
- [31] Sukhramani, P. S., Sukhramani, P. S., Desai, S. A., and Suthar, M. P. 2011. In-vitro Cytotoxicity Evaluation of Novel N-Substituted Bis-Benzimidazole Derivatives for Anti-Lung and Anti-Breast Cancer Activity. Annals of Biological Research. 2(1): 51-59.
- [32] Jung, G.-Y., Park, Y.-J., and Han, J.-S. 2010. Effects of HA Released Calcium Ion on Osteoblast Differentiation. Journal of Materials Science: Materials in Medicine. 21(5): 1649-1654.
- [33] Mahavorasirikul, W., Viyanant, V., Chaijaroenkul, W., Itharat, A., and Na-Bangchang, K. 2010. Cytotoxic Activity of Thai Medicinal Plants Against Human Cholangiocarcinoma, Laryngeal and Hepatocarcinoma Cells in Vitro. BMC Complementary and Alternative Medicine. 10(1): 55.

- [34] Park, J.-B. 2012. Effects of  $17-\alpha$  ethynyl estradiol on Proliferation, Differentiation & Mineralization of Osteoprecursor Cells. The Indian Journal of Medical Research. 136(3): 466-470.
- [35] Sugawara, Y., Suzuki, K., Koshikawa, M., Ando, M., and lida, J. 2002. Necessity of Enzymatic Activity of Alkaline Phosphatase for Mineralization of Osteoblastic Cells.
- The Japanese Journal of Pharmacology. 88(3): 262-269.
- [36] Heino, T. J., Hentunen, T. A., and Väänänen, H. K. 2004. Conditioned Medium from Osteocytes Stimulates the Proliferation of Bone Marrow Mesenchymal Stem Cells and Their Differentiation into Osteoblasts. Experimental Cell Research. 294(2): 458-468.